

Metstoich – Teaching Quantitative Metabolism and Energetics in Biochemical Engineering

Kelvin W. W. Wong and John P. Barford*

Chemical & Biomolecular Engineering, HKUST

Contact person: Prof. Barford

Phone: (852) 2358 7237

Fax: (852) 2358 0054

Address: Room 4552, Academic Building, The Hong Kong University of Science and Technology, Clear Water Bay, Kowloon, HONG KONG

Email: barford@ust.hk

Keywords: biochemical engineering, metabolic flux analysis, quantitative metabolism

Abstract

Metstoich, a metabolic calculator developed for teaching, can provide a novel way to teach quantitative metabolism to biochemical engineering students. It can also introduce biochemistry / life science students to the quantitative aspects of life science subjects they have studied. Metstoich links traditional biochemistry-based metabolic approaches with practical engineering parameters. This allows students to understand the relationship between the quantitative aspects of metabolism and practical engineering applications.

Introduction

Biochemistry is one of the important foundation courses in a biochemical engineering curriculum. It provides a basic introduction of cellular metabolism to engineering students in order to teach them how raw materials can be converted into valuable

* barford@ust.hk

metabolic products by microorganisms in various bioprocesses. Teaching metabolism in biochemical engineering courses normally adopts the traditional “biochemistry approach”. Students are presented with a number of reaction pathways which make major cell components (e.g. protein, RNA, DNA, lipids, cell walls) as well as major catabolic products using a qualitative description. However, traditional chemical engineering courses focus on product yield, selectivity, reaction rate and reactor / process design. It is similar for biochemical engineers that product yield, biomass yield, ATP yield are important parameters for bioreactor design. All these goals, if applied to a biochemical system, require a quantitative knowledge of metabolism. Therefore, a quantitative description in metabolism can complement the major skill base of engineering students and is more consistent with the overall philosophy or learning outcomes of an engineering degree.

As a sub-set of system biology, metabolic engineering focus on the metabolism of one organism. It is the practice of purposeful modification of metabolism using recombinant DNA technology with mathematical analysis to optimize genetic and regulatory processes within the cell. This leads to the modification the cell’s properties, in order to achieve a desirable objective.^[1-4] Metabolic Flux Analysis (MFA, also known as metabolite balancing, metabolic flux balancing, etc.), is a practical tool to understand and analyze metabolic pathways, pathway interaction and control. Varma and Palsson^[5] suggested that there are five major applications for MFA, namely: (1) to quantify metabolic physiology, (2) to simulate and interpret experimental data, (3) to analyze metabolic pathways for metabolic engineering, (4) to optimize cell culture medium, and (5) to design and optimize bioprocesses. MFA is an analytical tool developed based on stoichiometric network models^[6], and it is assumed that those metabolic fluxes are in steady state when compared with growth and other processes. Unlike simulations-based on mechanistic models that require detailed enzyme kinetic data, MFA is used to analyze the metabolic flux map and only requires metabolic reaction pathway details and stoichiometry, growth metabolism, and several strain-specific parameters. MFA determines a domain of stoichiometrically allowable flux distributions.^[5] Even if several restrictions are enforced, for complete metabolite balancing of a cell, a very large amount of flux data needs to be analyzed in order to accurately represent the interactions between the various metabolic pathways. Practically, such analysis is assisted by specifically

designed software packages that simulate the metabolic networks.

The analysis provided by MFA is also good for demonstration of the quantitative aspects of metabolism to students. However, most analytical software packages are developed for research purpose and mainly focus on pathway control (i.e. metabolic control analysis, MCA). Metstoich was initially developed to focus on teaching metabolism and to link practical biochemical engineering parameters with metabolic flux analysis.

Existing Software Packages

To explore the physiological properties of biological system, a system of equations must be solved. Such a task can be easily done with the aid of modern personal computers and metabolic engineering software packages. Some important / widely used software packages are:

GEPASI 3^[7-8] is a widely used free biochemical reactions simulation software package. GEPASI simulates the kinetics of biochemical reaction systems and provides functions such as metabolic control analysis (MCA), elementary mode analysis (EMA), optimisation and parameter fitting. The last version of GEPASI released was 3.30 at September, 2002. COPASI^[9] was developed based on GEPASI with different simulation techniques, optimization routine, etc. Jarnac (a.k.a. Scamp II)^[10] simulates the steady state and transient behaviour of metabolic pathways and calculates all coefficients for MCA. E-Cell^[11] is an object-oriented, whole-cell simulation software package. MIST^[12] performs dynamic simulations, stoichiometric calculations and MCA. JWS Online^[13] is an Internet-based metabolic simulator with collections of several metabolic models, and it can provide MCA to analyse the simulation results. KINSIM^[14-15] is a rate equation based numerical simulator and it was used for the simulation of enzymatic reaction system kinetics. FluxAnalyzer^[16] is a MATLAB package with GUI for stoichiometric analysis of metabolic networks. It can provide functions such as MFA, flux optimisation, topological features detection, and pathway analysis. In one of the more extensive

examples, Klamt, et al.^[17] carried out a metabolic flux analysis on Purple Nonsulfur Bacteria by using FluxAnalyzer. This model involved 30 of the most important catabolic branchpoint-metabolites (intermediate metabolites to which at least three reactions are linked) and 41 catabolic reactions – 1 for growth rate, 25 for central metabolic pathways, and 7 for photosynthesis, cyclic electron transport during photosynthesis, respiration, ATP synthesis and maintenance. The model also involved 46 anabolic reactions using the stoichiometries presented in Neidhardt, et al.^[18]

Except for FluxAnalyzer, all above simulation packages focus on the dynamic behaviour of metabolic pathways. They require reaction kinetics as input and some of them can even perform metabolic pathway analysis such as MCA, etc.

Other than above listed software packages, there are many packages / projects developed / under development. Figure 1 summarizes part of metabolic engineering software packages / projects found. Most of them are ODE solvers, some of which can perform sensitivity analysis (MCA). However, some they were developed for various purposes, such as:

- CellDesigner^[19] is for gene-regulatory and biochemical networks.
- Cellerator^[20] is a Mathematica package designed for modeling with automated equation generation. It was designed with the intent of simulating signal transduction.
- InNetics^[21] was developed for genomic-based drug discovery.
- The JigCell project^[22] is to explore the cell physiology from the scope of molecular regulatory networks.

There are two trends for metabolic software development. The first trend is using visual tools to allow users to construct pathway models. The second is the development of web based applications. Nowadays, the Internet is already part of our daily life and web based applications are a good choice, especially for database projects to collect and share data.

The common advantage of these packages is that you can input any model to the

package for analysis. However, their practical use for engineering purposes is limited and not their primary purpose. They do not address issue of energetics and ATP usage, the production of biomass yield, etc.

Metstoich

Metstoich was initially developed for teaching purposes^[23-24] and is based on the metabolism of a specific yeast, *S. cerevisiae*^[25]. Metstoich includes the following major pathways: (1) central metabolic pathways, such as glycolysis, tricarboxylic acid (TCA) cycle and pentose-phosphate pathway (PPP), and (2) biosynthetic pathways. The central metabolic pathways serve to provide precursors for biosynthetic pathways, and for generating energy (ATP) to support cell growth and maintenance.

The main purpose of Metstoich is to link metabolic flux distribution among pathways with practical engineering parameters encountered in standard biochemical engineering course, such as biomass yield (Y_{XS}), product yield (Y_{PS}), ATP yield ($Y_{X/ATP}$), etc. Pathway reactions are predefined and based on a specific yeast. Such approach could also help to identify flux distribution among branch points.

There are several important inputs necessary for Metstoich to determine the flux map:

- (1) Cell macromolecular composition;
- (2) Glucose distribution (usage) in central metabolic pathways for energy generation process;
- (3) P/O ratio;
- (4) ATP utilization efficiency (or simply called as ATP efficiency, η), that is the percentage of total ATP generated that is directly consumed in biosynthetic reactions;
- (5) Biomass yield, Y_{XS} ;
- (6) ATP yield, $Y_{X/ATP}$.

There are four problem types which can be solved by Metstoich with above inputs:

- (a) Calculation based on theoretical yield; or

- (b) Calculation based on experimental biomass yield, Y_{XS} ; or
- (c) Calculation based on predefined ATP yield, $Y_{X/ATP}$; or
- (d) Calculation based on experimental biomass yield, Y_{XS} , and predefined ATP yield, $Y_{X/ATP}$.

Table 1 summarizes matrix of problem types, inputs and outputs and Figure 2 shows part of the input interface.

Users can specify the cell composition (Figure 2), carbon source and electron donor if CO₂ is the carbon source (Figure 3), electron acceptor (Figure 4) and energetic issues of the microorganism (Figure 5). Metstoich can compare two sets of metabolic flux maps (Figure 6) and highlights fluxes with a defined degree of difference in percentage.

The results generated by Metstoich are organized into several levels of detailed worksheets with biochemical detail and illustrative reaction pathways included to make it more understandable. Levels of organized results are:

Cell Yield and Energetics (Figure 7) – This worksheet is the executive summary of the overall performance of the cell with the inputted common engineering parameters;

Fate of Glucose (Figure 8) – This worksheet summarizes how much glucose is used for specified purposes via specified pathways;

Composition Summary (Figure 9) – This worksheet summarizes cell compositions and their detail; and

All Detailed Reactions (Figure 10) – This worksheet shows the detailed flux maps for biosynthetic pathways, central metabolic pathways used for either biosynthesis purpose or energy generation purposes.

Metstoich already contains amino acid production pathways and it is capable to analyze amino acid production. Since Metstoich already contains information on major catabolic and anabolic pathways. It can be easy to further include more production formation pathways such as antibodies, biofuel, etc.

Since Metstoich is focused on the static metabolic flux analysis, enzyme concentrations, kinetic expressions, intermediate concentrations, and thermodynamics

have not been incorporated. An extension of Metstoich which incorporate thermodynamics, reaction kinetics, etc. has been developed and reported.^[26-28]

The core calculation module of Metstoich is written using Microsoft Excel 2002 with VBA Macro. This core Excel module is responsible for constructing and displaying the metabolic flux map. The front-end graphical user interface was written in Visual Basic. Metstoich runs on Microsoft Windows 98, 2000, XP and Vista with Microsoft Office 2000, XP or 2003 installed.

Example to Demonstrate the Teaching of Quantitative Metabolism to Students

This is an example problem that students to undertake as an exercise. It is taken from a number of problems included in the Metstoich package:

The biomass composition (weight %) of a given yeast is as follows:

Protein = 39%, DNA = 1%, RNA = 11%, Lipids = 3%, Phospholipids = 5%,
Cell Wall = 38%, and Ash = 3%

For energy generation, 10% glucose is used by pentose phosphate pathway, 60% glucose is used by the TCA cycle and 30% glucose is used by the fermentation pathway. The reported biomass yield is 0.4 g-biomass / g-glucose and let P/O ratio as 2.2 mol-ATP / mol-NADH. What is the corresponding $Y_{X/ATP}$ and ATP efficiency. What is the relationship in between P/O ratio and $Y_{X/ATP}$?

Since Y_{XS} with P/O ratio are given, the “Experimental Y_{XS} ” calculation mode should be used. With given input values, Metstoich returns $Y_{X/ATP} = 7.85$ g-biomass / mol-ATP and ATP efficiency is 10.6%. And the relationship between $Y_{X/ATP}$ and P/O ratio is shown in Figure 11 at various P/O ratios:

With fixed Y_{XS} and cell compositions, glucose directly consumed to form biomass is always fixed at 1.43 g-glucose / g-biomass. The total glucose consumed is 2.5 g-glucose / g-biomass for the given $Y_{XS} = 0.4$. Therefore glucose consumed to generate energy is always 1.07 g-glucose / g-biomass, and it always generates 17.3 mmol-ATP and 50.1 mmol-NADH per 1.07 g-glucose consumed in assigned

pathways. Therefore, total ATP generated in energy generation process = $(17.3 + P/O \times 50.1 \text{ mmol-ATP}) / 1.07 \text{ g-glucose}$. And $Y_{X/ATP} = 1 \text{ g-biomass} / \text{total ATP}$ generated in energy generation process. It is suggested that normal $Y_{X/ATP}$ is around $10.5 \text{ g-biomass} / \text{mol-ATP}$. With using the “Experimental Y_{XS} and Fixed $Y_{X/ATP}$ ” calculation mode, it is found that the P/O ratio = $1.56 \text{ mol-ATP} / \text{mol-NADH}$ and ATP efficiency = 14.27%.

Based on the fluxes given by Metstoich, students can draw simplified flux map as illustrated in Figure 12, Figure 13 and Figure 14 to understand the quantitative use of glucose by the cell and how much energy had been generated. By combining Figure 13 and Figure 14, students can generate an overall quantitative flux distribution for the given biomass.

Comments on Metstoich

Professional evaluation was undertaken by Learnet of Hong Kong University. Metstoich had been reviewed by 4 leading academics in biochemical engineering from UK, USA, Australia and Singapore: Prof. D. Bogle from University College London, Prof. L. Nielsen from University of Queensland, Prof. D. Trau from National University of Singapore and Prof. P. Fu from University of Hawaii at Manoa. It was considered an excellent tool for learning of major biochemical engineering concepts such as $Y_{X/ATP}$, yield, etc. The feature which compared two sets of metabolic flux maps with a percentage change larger than a specified number was also highly regarded (Figure 6). In general, Metstoich has been rated as 4 stars out of 5 by these academics for different aspects such as interface design, quality of content, and learning potential.

Metstoich as Teaching Tool

Metstoich had been applied in biochemical engineering and biochemistry classes in HKUST and it has been rated as easy to use by students. Students had been

interviewed by the Center of Enhanced Learning & Teaching (CELT) of the Hong Kong University of Science and Technology (HKUST). It is agreed that Metstoich is easy to use, since the help functions and labels and button of the software are clear. The advantage of Metstoich is: it can compare two sets of calculated results by highlighting the difference. However, students feel that Metstoich contain too much information since it covers from networks of reactions to energetics and cell yield, etc.

Conclusion

Engineering students are used to quantitative concepts from their foundation courses. Biochemistry can also be taught quantitatively and when this is done, engineering students can appreciate the importance of metabolism in understanding and optimising bioprocesses. Metstoich, a metabolic calculator for teaching purposes, was developed to introduce metabolism to students using quantitative principles. As such it is useful to both engineering students and biochemistry / life sciences students, who normally do not have strong background or training in quantitative methods.

Metstoich has many unique / novel features:

1. Linking practical engineering parameters with cell growth, product yield, energetics, etc.
2. Analysing the flux through any reaction pathway;
3. Calculating how much nutrients are required for cell growth.

Such analysis can provide useful information about how product yield is related with biomass yield, cell energetics, etc. Students can explore different metabolic options and are challenged to further explore their relationship to bioreactor / medium design.

The package has been well received by both academic experts in biochemical engineering and undergraduate chemical engineering and biochemistry students at HKUST.

Acknowledgement

The authors would like to acknowledge the financial support (Project : HKUST 00409E) of the Center For Enhanced Learning & Teaching (CELT) as well as their technical participation in the project.

References

1. Nielsen, J., "Metabolic Engineering: Techniques for Analysis of Targets for Genetic Manipulations", *Biotech. Bioeng.*, **58**, 125-132 (1998)
2. Olsson, L., and J. Nielsen, "The Role of Metabolic Engineering in the Improvement of *Saccharomyces cerevisiae*: Utilisation of Industrial Media", *Enzyme Microb. Technol.*, **26**, 785-792 (2002)
3. Stephanopoulos, G. N., A. A. Aristidou and J. Nielsen, *Metabolic Engineering, Principles and Methodologies*, Academic Press, (1998)
4. Lee, S. Y., and E. T. Papoutsakis Ed., *Metabolic Engineering*, Marcel Dekker, Inc., (1999)
5. Varma, A., and B. O. Palsson, "Metabolic Flux Balancing: Basic Concepts, Scientific and Practical Use", *Bio/technology*, **12**, 994-998 (1994)
6. Weichert, W., "Modeling and Simulation: Tools for Metabolic Engineering", *J. Biotech.*, **94**, 37-63 (2002)
7. Mendes, P., "Biochemistry by Numbers: Simulation of Biochemical Pathways with Gepasi 3", *Trends Biochem. Sci.*, **22**, 361-363 (1997)
8. Mendes, P., <http://www.gepasi.org/>
9. COPASI.org, <http://www.copasi.org/>
10. Sauro, H. M., "Scamp: A General-Purpose Simulator and Metabolic Control Analysis Program", *Comput. Appl. Biosci.*, **9**, 441-450 (1993)
11. Tomita, M., K. Hashimoto, K. Takahashi, T. S. Shimizu, Y. Matsuzaki, F. Muiyoshi, K. Saito, S. Tanida, K. Yugi, J. C. Venter, and C. A. Hutchison, 3rd, "E-CELL: Software Environment for Whole-Cell Simulation", *Bioinformatics*, **15**, 72-84 (1999)

12. Ehilde, M., and G. Zacchi, "Mist: A User-friendly Metabolic Simulator", *Comp. App. Biosci. (CABIOS)*, **11**, 201-207 (1995)
13. Olivier, B., and J. Snoep, <http://jjj.biochem.sun.ac.za/>, (2002-2003)
14. Barshop, B. A., R. F. Wrenn and C. Frieden, "Analysis of Numerical Methods for Computer Simulation of Kinetic Processes: Development of KINSIM – A Flexible, Portable system", *Anal. Biochem.*, **130**, 134-145 (1983)
15. Dang, Q., and C. Frieden, "New PC Versions of the Kinetic-Simulation and Fitting Programs, KINSIM and FITSIM", *Trends Biochem. Sci.*, **22**, 317 (1997)
16. Klamt, S., J. Stelling, M. Ginkel, and E. D. Gilles, "FluxAnalyzer: Exploring Structure, Pathways, and Flux Distributions in Metabolic Networks on Interactive Flux Maps", *Bioinformatics*, **19**, 261-269 (2003)
17. Klamt, S., S. Schuster, and E. D. Gilles, "Calculability Analysis in Underdetermined Metabolic Networks Illustrated by a Model of the Central Metabolism in Purple Nonsulfur Bacteria", *Biotech. Bioeng.*, **77**, 734-751 (2002)
18. Neidhardt, F. C., J. L. Ingraham, and M. Schaechter, *Physiology of the Bacterial Cell – A Molecular Approach*, Sinauer Associates, (1990)
19. CellDesigner.org, <http://celldesigner.org/>
20. Cellerator.info, <http://www.cellerator.info/>
21. InNetics.com, <http://innetics.com/>
22. JigCell Project, <http://jigcell.biol.vt.edu/>
23. Wong, K. W., J. P. Barford and J. F. Porter, "Understanding the Practical Consequences of Metabolic Interactions – A Software Package for Teaching and Research", *Computer Applications in Biotechnology 9th International Symposium*, Nancy, France, (2004)
24. Wong, K. W., J. P. Barford and J. F. Porter, "Understanding the Practical Consequences of Metabolic Interactions – A Software Package for Teaching and Research", *Proceedings of the Second Teaching & Learning Symposium*, The Hong Kong University of Science & Technology, Hong Kong, (2004)
25. Oura, E., "The Effect of Aeration on the Growth Energetics and Biochemical Composition of Baker's Yeast" with appendix: "Reactions Leading to the Formation of Yeast Cell Material from Glucose and Ethanol", PhD Thesis, Helsinki University, Helsinki, Finland, (1972)
26. Sanderson, C.S., Barford, J.P. and Barton, G.W., "A structured dynamic model for animal cell culture systems", *Biochem. Eng. Journal*, **3**, 203-211 (1999)

27. Sanderson, C.S., Barford, J.P., Barton, G.W., Wong, T.K. and Reid, S, “A structured dynamic model for animal cell culture systems: application to baculovirus / insect cell systems”, *Biochem. Eng. Journal*, **3**, 219-229, (1999)
28. Sanderson, C.S., Jang, J.D., Barford, J.P. and Barton, G.W., “A structured dynamic model for animal cell culture systems: application to murine hybridoma”, *Biochem. Eng. Journal*, **3**, 213-218 (1999)

Figure Captions

[Figure 1](#) – Some Existing Metabolic Engineering Software Packages / Projects.

[Figure 2](#) – Part of Metstoich Input Interface – Basic Information and Cell Compositions

[Figure 3](#) – Part of Metstoich Input Interface – Carbon Source

[Figure 4](#) – Part of Metstoich Input Interface – Electron Acceptors

[Figure 5](#) – Part of Metstoich Input Interface – Energetic and Other Parameters

[Figure 6](#) – User Can Specify Highlight Values That Changed Larger than the Given Percentage,

[Figure 7](#) – “Cell Yield and Energetics”, the cell yield (either estimated or given), $Y_{X/ATP}$, amount of ATP generated directly from reactions or oxidative phosphorylation are summarized.

[Figure 8](#) – “Fate of Glucose”, glucose directly linked with biosynthesis or energy generation process is analyzed.

[Figure 9](#) – All detailed biomass compositions, such as amino acid, etc. are summarized in “composition summary”.

[Figure 10](#) – “All Detailed Reactions” shows all biochemical reactions.

[Figure 11](#) – Relationship in between $Y_{X/ATP}$ and P/O ratio.

[Figure 12](#) – Glucose used for biosynthesis and energy generation purposes, drawn based on Metstoich results.

[Figure 13](#) – Fluxes among central metabolic pathways for biosynthesis, drawn based

on Metstoich results.

[Figure 14](#) – Fluxes among central metabolic pathways for energy generation purpose, drawn based on Metstoich results.

Table

Parameters	Problem Types			
	(a) Theoretical Y_{XS}	(b) Experimental Y_{XS}	(c) Predefined $Y_{X/ATP}$	(d) Experimental Y_{XS} with Predefined $Y_{X/ATP}$
(1) Cell compositions	Input	Input	Input	Input
(2) Glucose usage for energy generation process	Input	Input	Input	Input
(3) P/O ratio	Input	Input	Output	Output
(4) ATP efficiency	Input	Output	Input	Output
(5) Y_{XS}	Output	Input	Output	Input
(6) $Y_{X/ATP}$	Output	Output	Input	Input

Table 1 – Parameters as Input & Output for Various Calculation Modes in Metstoich